

Technical Training :Xpert[®] Xpress SARS-CoV-2

For use with GeneXpert[®] Systems with Touchscreen



Cepheid.

GeneXpert.

In Vitro Diagnostic Medical Device

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Training Agenda *Xpert® Xpress SARS-CoV-2*

1	Reagents	
2	Kit Storage and Handling	GeneXpert.
3	Specimen Collection, Storage, and Handling	Xpert® Xpress SARS-CoV-2
4	Preparing the Cartridge	303456/7890 123456/ 123456/
5	Quality Controls	Far use under Emergency Use Authorization (EUA)
6	Results Analysis	
7	Discussion	



Training Objectives

At the end of the training, users will be able to:

- Properly store and handle the Xpert® Xpress SARS-CoV-2 kit
- Follow proper laboratory safety precautions
- Collect and store appropriate specimen(s)
- Prepare a cartridge and run the Xpert® Xpress SARS-CoV-2 test
- Report the various software generated results
- Understand the Xpert[®] Xpress SARS-CoV-2 control strategy





The Cepheid Solution



Detection of SARS-CoV-2

- On-board internal controls for each sample
 - Probe Check Control (PCC)
 - Sample Processing Control (SPC)
- Closed cartridge system minimizes risk of contamination
- On-demand results
- Random access



Intended Use

- The Xpert[®] Xpress SARS-CoV-2 test is a real-time RT-PCR test intended for the qualitative detection of nucleic acid from the SARS-CoV-2 in nasopharyngeal swab, nasal swab, or nasal wash/aspirate specimen collected from individuals who are suspected of COVID-19 infection.
- Results are for the identification of SARS-CoV-2 RNA. Positive results are indicative of the
 presence of SARS-CoV-2 RNA; clinical correlation with patient history and other diagnostic
 information is necessary to determine patient infection status. Positive results do not rule
 out bacterial infection or co-infection with other viruses. The agent detected may not be the
 definite cause of disease.
- Negative results do not preclude SARS-CoV-2 infection and should not be used as the sole basis for treatment or other patient management decisions. Negative results must be combined with clinical observations, patient history, and epidemiological information.
- The Xpert[®] Xpress SARS-CoV-2 test is intended to be performed by trained users in both laboratory and near patient testing settings.



Good Laboratory Practice Review

Personnel Protective Equipment (PPE)

- Wear clean lab coats, safety glasses, and gloves
- Change gloves between processing samples

Lab Bench Area

- Clean work surfaces routinely with:
 - 1:10 dilution of household bleach*
 - ✓ 70% ethanol solution
- After cleaning, ensure work surfaces are dry

- Use filtered pipette tips when recommended
 - Follow the manufacturer's requirements for calibration and maintenance of equipment

Equipment





 Store specimens and sample away from kit to prevent contamination

Specimens, Samples, and Kits Storage

*Final Active Chlorine concentration should be 0.5% regardless of the household bleach concentration in your country.

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Kit Storage and Handling

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Xpert® Xpress SARS-CoV-2 Requirements

GeneXpert® Systems

 For GeneXpert[®] System with Touchscreen: Cepheid OS 1.0



Test Kits

XPRSARS-COV2-10

Materials Required by Not Provided

- Nylon flocked swab (Copan P/N 502CS01, 503CS01) or equivalent
- Viral transport medium, 3 mL (Copan P/N 330C) or equivalent
- 0.9% (w/v) saline, 3 mL
- Sample Collection Kit for Viruses (Copan P/N 305C, Copan P/N 346C) or equivalent
- Personal Protective Equipment (PPE)
- 1:10 Bleach
- 70% ethanol or denatured ethanol

Optional

- Uninterruptible Power Supply/Surge Protector
- Printer



Xpert[®] Xpress SARS-CoV-2 *Kit Components*

Catalog Number	XPRSARS-COV2-10
Tests per Kit	10
	Assay Definition File (ADF)
Kit CD	Assay Import Instructions
	Flyer: Instructions to access on-line reference materials including the Product Insert
Disposal Transfer Pipettes	10 to 12
Storage	2–28°C



Cartridges contain chemically hazardous substances. Please see Instructions for Use and Safety Data Sheet for more detailed information.



Xpert[®] Xpress SARS-CoV-2 *Kit Storage and Handling*

- Store Xpert[®] Xpress SARS-CoV-2 cartridges and reagents at 2–28°C.
- Follow your institution's safety procedures for working with chemicals and handling biological samples
- Do not use collection devices that have not been validated by Cepheid
- Open the cartridge lid only when adding the sample, close the lid, and proceed with processing
 - Start the test within 30 minutes of adding the sample to the cartridge.



Warnings and Precautions



Do not shake the cartridge

- Do not use a cartridge if it...
 - appears wet, has leaked, or if the lid seal appears to have been broken
 - appears damaged
 - has been dropped after removing it from packaging
 - has been dropped or shaken after you have added the sample
 - has a damaged reaction tube
 - has been used; each cartridge is single-use to process one test
 - has expired

Do not reuse pipettes

Dispose of cartridges and reagents according to your institution's and country's guidelines for disposal of hazardous materials



Warnings and Precautions

Biological specimens, transfer devices, and used cartridges should be considered capable of transmitting infectious agents and require use of standard precautions.

Follow your institution's environmental waste procedures for proper disposal of used cartridges and unused reagents.

These materials may exhibit characteristics of chemical hazardous waste requiring specific national or regional disposal procedures.

If national or regional regulations do not provide clear direction on proper disposal, the biological specimens and used cartridges should be disposed of per WHO (World Health Organization) medical waste handling and disposal guidelines.





Limitations

- Performance characteristics of this test have been established with the specimen types listed in the Intended Use Section only. The performance of this assay with other specimen types or samples has not been evaluated.
- A false negative result may occur if a specimen is improperly collected, transported or handled. False negative results may also occur if inadequate numbers of organisms are present in the specimen.
- As with any molecular test, mutations within the target regions of Xpert[®] Xpress SARS-CoV-2 could affect primer and/or probe binding resulting in failure to detect the presence of virus.
- This test cannot rule out diseases caused by other bacterial or viral pathogens.



For detailed information, refer to the current Instructions For Use.



Specimen Collection, Storage and Handling

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Specimen Collection

Specimen Type: Nasopharyngeal swab nasal swab, and/or nasal wash/aspirate specimens

Place specimen into 3m transport medium or 3mL of saline



Refer to the WHO Laboratory Biosafety Guidance Related to the Coronovirus Disease 2019 (COVID-19):

https://www.who.int/publications-detail/laboratory-biosafetyguidance-related-to-coronavirus-disease-2019-(covid-19)



Specimen Collection: Nasopharyngeal Swab

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Insert the swab into either nostril, passing it into the posterior nasopharynx.

Ζ	

Rotate swab by firmly brushing against the nasopharynx several times.

3

Remove and place the swab into the tube containing 3mL of viral transport medium or 3mL of saline.



Break swab at the indicated break line and cap the specimen collection tube tightly.





Specimen Collection: Nasopharyngeal Swab

Nasopharyngeal Specimen Collection

For use with Xpert® Nasopharyngeal Sample Collection Kit - Catalog # SWAB/B-100

Open the package that contains the swab and transport medium tube. Set the tube aside before collecting the specimen.

2



Remove the cap from the tube. Insert the swab into the transport medium.





Break the swab shaft against the side of the tube at the scoreline. 6

5

Avoid splashing contents on the skin. Wash with soap and water if exposed.







For Xpert Xpress SARS-CoV-2: Specimen may be stored for 8 hours at 15-30°C or up to 7 days at 2-8°C.











In Vitro Diagnostic Use ((IVD



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Specimen Collection: Nasal Swab



Insert the nasal swab 1 to 1.5cm into the nostril.



Rotate the swab against the inside of the nostril for 3 seconds while applying pressure with a finger to the outside of the nostril.



Repeat on the other nostril with the same swab.



Remove and place the swab into the tube containing 3mL of viral transport medium or 3mL of saline.



Break swab at the indicated break line and cap the specimen collection tube tightly.





Specimen Collection: **Nasal Swab**



For use with Xpert® Swab Sample Collection Kit - Catalog # SWAB/F-100

Open the package that contains the swab and transport medium tube. Set the tube aside before collecting the specimen.



- Repeat Step 4 on the other nostril with the same swab.
- To avoid specimen contamination, do not touch the swab tip to anything after collecting the specimen.





In Vitro Diagnostic Use CE IVD

Remove the cap from the tube. Insert the swab into the transport medium.

Break the swab shaft against the side of the tube at the scoreline.

Avoid splashing contents on the skin. Wash with soap and water if exposed.

Replace the cap on the tube and close tightly.

For Xpert Xpress Flu and Xpert Xpress Flu/RSV: Specimen may be stored for 24 hours at 15-30°C or up to 7 days at 2-8°C.

For Xpert Xpress SARS-CoV-2: Specimen may be stored for 8 hours at 15-30°C or up to 7 days at 2-8°C.





the swab, taking care not to touch the tip of the swab to any surface.

Hold the swab in your hand, pinching

in the middle of the swab shaft on the

Rotate swab against the inside of the nostril for 3 seconds while applying

pressure with a finger to the outside of

* SWAB/F-100 contains Copan UTM 330C and Copan nylon swab 502CS01

Do not insert the swabs more than

2

3

4

scoreline.

the nostril.

1-1.5 cm.

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Specimen Collection: Nasal Wash/Aspirate

Nasal wash/aspirate specimens can be collected following the user institution standard procedure. Also, refer to the WHO guidelines for the collection of human nasal wash/aspirate specimens.

- 1. Using a transfer pipette, transfer 600µL of the undiluted nasal wash/aspirate specimen into the tube containing 3mL of viral transport medium or 3mL of saline.
- 2. Cap the tube.

https://www.who.int/influenza/human_animal_interface/virology_laboratories_ and_vaccines/guidelines_collection_h5n1_humans/en/



Specimen Transport and Storage

Sample Type

Viral Transport Medium or saline containing:

Nasopharyngeal swab

OR

Nasal swab

OR

Nasal wash/aspirate specimens

Transport and Storage Conditions







Cartridge Preparation

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Proper Cartridge Handling Techniques



• Do not touch the reaction tube

Correct

- Keep the cartridge upright after seal has been broken
- Do not tilt when scanning the cartridge





Xpert[®] Xpress SARS-CoV-2 Cartridge Preparation



Sample Qualification—Check if all items below are present:

- 1. Transport media containing swab (if applicable)
- 2. Patient name or identifier on the tube
- 3. Cartridges and transport media are within the expiration date

Good Laboratory Practices

- · Wear clean gloves and lab coats
- Change gloves between samples
- Clean work surface with 1:10 dilution of bleach followed by 70% ethanol solution



Xpert[®] Xpress SARS-CoV-2 Cartridge Preparation





Run a Test Start the Test Within 30 Minutes





For complete details on how to run a test, refer to the Instructions For Use and the GeneXpert[®] System with Touchscreen Operator Manual.

Automated Xpert® Xpress SARS-CoV-2 Protocol



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Quality Controls

Refer to the Instructions For Use for complete details

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Assay Control Strategy

Xpert® Xpress SARS-CoV-2 Quality Controls

- Each Xpert[®] cartridge is a self-contained test device
- Cepheid designed specific molecular methods to include internal controls that enable the system to detect specific failure modes within each cartridge*
 - 1. Sample Processing Control (SPC)
 - 2. Probe Check Controls (PCC)



*Refer to 301-4868 GeneXpert® Quality Control Features for All Cepheid Xpert® Assays



Internal Quality Controls

Probe Check Controls (PCC)

- Before the PCR step, fluorescence signal is measured on all probes and compared with default factory settings to monitor
 - Reagent rehydration
 Probe integrity

- PCR tube filling
- Dye stability

Sample Processing Controls (SPC)

- Non-infectious spore in each cartridge
 - Verifies adequate sample processing
 - Verifies lysis and detects PCR inhibition
 - Should be positive in a negative sample
 - Can be positive or negative in a positive sample



Commercially Available External Controls



To minimize degradation of the control material, return any unused sample to the recommended storage conditions immediately after use.

- Many other vendors for quality control material are also available in addition to the one outlined above.
- External controls should be used in accordance with local, state accrediting organizations, as applicable



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Result Interpretation

Refer to the Instructions For Use for complete details

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Early Assay Termination

- The Xpress SARS-CoV-2 test includes an Early Assay Termination (EAT) function that will provide earlier time to result in high titer specimens.
- When SARS-CoV-2 titers are high enough to initiate the EAT function, the SPC amplification curve may not be seen, and its results may not be reported.



Results Summary

Result Displayed	N2	E	SPC
	+	+	- L /
SANS-COV-Z FOSITIVE	+	_	T /
SARS-CoV-2 PRESUMPTIVE POS	_	+	+/
SARS-CoV-2 NEGATIVE	_	_	+
INVALID	_	_	_
ERROR	NO RESULT	NO RESULT	NO RESULT
NO RESULT	NO RESULT	NO RESULT	NO RESULT



SARS-CoV-2 POSITIVE

Result SARS-CoV-2 POSITIF

«BACK HOME **RESULTS** QC ADMIN **Test Completed** REPORT Module B2 Result SARS-CoV-2 POSITIE BCC38BFA5CE90094CD584D8 Sample ID 47 Patient ID Specimen Test Type Xpert Xpress SARS-CoV-2 Assay Name User cepheid Start Date & Time 06/05/20 16:04:46 **Test Disclaimer** For In Vitro Diagnostic Use Name Only. For use under the **Emergency Use Authorization** Е (US). N2 SPC

Test Report Patient ID: Sample ID: BCC38BFA5CE90094CD584D847 Test Type: Specimen Assay Information Assay Name Assay Version Assay Type Xpert Xpress SARS-CoV-2 2 In Vitro Diagnostic SARS-CoV-2 POSITIF Test Result: Analyte Result Analyte Ct EndPt Analyte Probe Result Check Result 40.9 47 POS PASS 38.8 174 POS PASS 27.8 342 NA PASS

The 2019 novel coronavirus (SARS-CoV-2) target nucleic acids are detected.

- The SARS-CoV-2 signal for the N2 nucleic acid target or signals for both nucleic acid targets (N2 and E) have a Ct within the valid range and endpoint above the minimum setting
- SPC: NA; SPC is ignored because coronavirus target amplification occurred
- Probe Check: PASS; all probe check results pass



SARS-CoV-2 PRESUMPTIVE POS

Result SARS-CoV-2 PRESUMPTIVE POS

« BACK		HOME	RESULTS	QC	ADMIN	€
Test Comple	eted				REPORT	
Module L2		Result		_		
Sample ID	1010603573A	SARS-COV-2 PRES	SUMPTIVE POS			
Patient ID						
Test Type	Specimen					
Assay Name	Xpert Xpress SARS-CoV-2					
User	Nwe Tun					
Start Date & Time	04/15/20 23:32:21					
Test Disclaimer	For In Vitro Diagnostic Use Only. For use under the Emergency Use Authorization (EUA).					

			Test	Report	
Patient ID Sample II Test Type): D*:):	10106 Speci	03573A men		
Assay Info	ormation				
Assay Na	me			Assay Version	Assay Type
Xpert Xpres	s SARS-Co	V-2		1	In Vitro Diagnostic
lest Resu	ult:	SARS-CoV-	-2 PRESUM	PTIVE POS	
Analyte R	ult: Xesult	SARS-CoV-	-2 PRESUM	PTIVE POS	
Analyte R	uit: tesult Ct	SARS-CoV-	-2 PRESUM	PTIVE POS	
Analyte R Analyte R Analyte Name	uit: Result Ct	SARS-Cov-	Analyte Result	PTIVE POS Probe Check	
Analyte R Analyte R Analyte Name	ult: Xesult Ct	SARS-Cov-	Analyte Result	PTIVE POS Probe Check Result	
Analyte R Analyte R Analyte Name E	alt: Ct 39.2	EndPt	Analyte Result POS	Probe Check Result PASS	
Analyte R Analyte R Analyte Name E N2	ut: Ct 39.2 0.0	EndPt	Analyte Result POS NEG	Probe Check Result PASS PASS	

The 2019 novel coronavirus (SARS-CoV-2) nucleic acids may be present.

- Sample should be retested. For samples with a repeated Presumptive Positive result, additional confirmatory testing may be conducted, if it is necessary to differentiate between SARS-CoV-2 and SARS-CoV-1 or other Sarbecovirus currently unknown to infect humans, for epidemiological purposes or clinical management.
- The SARS-CoV-2 signal for only the E nucleic acid target has a Ct within the valid range and endpoint above the minimum setting
- SPC: NA; SPC is ignored because a target amplification has occurred.
- Probe Check: PASS; all probe check results pass



SARS-CoV-2 NEGATIVE

Result SARS-CoV-2 NEGATIVE

BACK			HOME	RESULTS	QC	ADMIN	
Test	Comple	ted				REPORT	
Modul	e D4		Result SARS-CoV-2 NEG/	ATIVE			
Sample	ID	Flu A-Flu B					
Patient	ID						
Test Ty	pe	Specimen					
Assay N	lame	Xpert Xpress_SARS-CoV-2					
User		JoAnn Kop					
Start Da	ate & Time	11/18/20 09:03:26					
Test Dis	sclaimer	For In Vitro Diagnostic Use Only. For use under the Emergency Use Authorization (US).					

			Test	Report	
Patient ID Sample II Test Type	: D: :	Flu A-F Specim	ilu B nen		
Assay Info	ormation				
Assav Na	me			Assay Version	Assay Type
					1 1 1
Xpert Xpres	s_SARS-Co	V-2		4	In Vitro Diagnostic
Xpert Xpres Test Resi Analyte F	s_SARS-Co ult: Result	V-2 SARS-CoV-2	2 NEGATIV	4 (E	In Vitro Diagnostic
Xpert Xpres Test Resi Analyte F Analyte	s_SARS-Co ult: Result Ct	V-2 SARS-CoV-2 EndPt	2 NEGATIV Analyte	4 /E Probe	In Vitro Diagnostic
Xpert Xpres Test Reso Analyte F Analyte Name	s_SARS-Co ult: Result Ct	V-2 SARS-CoV-2 EndPt	2 NEGATIV Analyte Result	4 /E Probe Check	In Vitro Diagnostic
Xpert Xpres Test Resu Analyte R Analyte Name	s_SARS-Co ult: Result Ct	V-2 SARS-CoV-2 EndPt	2 NEGATIV Analyte Result	4 /E Probe Check Result	In Vitro Diagnostic
Xpert Xpres Test Resu Analyte F Analyte Name SARS-CoV	s_SARS-Co ult: Result Ct -2 0.0	V-2 SARS-CoV-2 EndPt 2	2 NEGATIV Analyte Result NEG	4 /E Probe Check Result PASS	In Vitro Diagnostic

The 2019 novel coronavirus (SARS-CoV-2) target nucleic acids are not detected.

- The SARS-CoV-2 signals for two nucleic acid targets (N2 and E) do not have a Ct within the valid range and endpoint above the minimum setting
- SPC: PASS; SPC has a Ct within the valid range and endpoint above the minimum setting
- Probe Check: PASS; all probe check results pass





Troubleshooting

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Factors That Negatively Affect Results

- Improper specimen collection
 - The performance of this assay with other specimen types or samples has not been evaluated
- Inadequate numbers of organisms are present in the specimen
- Improper transport or storage of collected specimen
 - Storage and transport conditions are specimen specific
 - Refer to the Instructions For Use for the appropriate handling instructions
- Improper testing procedure
 - Modification to the testing procedures may alter the performance of the test
 - Careful compliance with the Instructions For Use is necessary to avoid erroneous results



NO RESULT - REPEAT TEST

ACK		HOME	RESULTS	QC	ADMIN	E
Test Comple	eted				REPORT	
Module D4		Result NO RESULT - REP	EAT TEST	_		
Sample ID	Test 01					
Patient ID						
Test Type	Specimen					
Assay Name	Xpert Xpress_SARS-CoV- 2_Flu_RSV					
User	Jun Zhang					
Start Date & Time	11/19/20 17:46:01					
Test Disclaimer	For In Vitro Diagnostic Use Only. For use under the Emergency Use Authorization (US).					

SPC does not meet acceptance criteria. Presence or absence of the target RNAs cannot be determined.

- SPC: FAIL;
- SARS-CoV-2, Flu A, Flu B, RSV signals do not have a Ct within valid range and endpoint below minimum setting
- Probe Check: PASS; all probe check results pass

			Test	Report	
Patient ID Sample ID):):	Test 0)1		
Test Type	:	Speci	men		
Assay Info	ormation				
Assay Na	me			Assay Version	Assay Type
Xpert Xpres	s_SARS-Co	v-2_Flu_RSV		4	In Vitro Diagnostic
Test Desi	.14.			TFOT	
Test Resu Analyte R	ult: Result	NO RESUL	T - REPEAT	TEST	
Test Resu Analyte R Analyte	ult: Result Ct	NO RESUL	T - REPEAT	Probe	
Test Resu Analyte R Analyte Name	ult: Result Ct	NO RESUL	T - REPEAT Analyte Result	TTEST Probe Check Result	
Test Resu Analyte R Analyte Name SARS-CoV-	ult: Result Ct -2 0.0	NO RESUL	T - REPEAT Analyte Result	TEST Probe Check Result PASS	
Test Resu Analyte R Analyte Name SARS-CoV- Flu A 1	ult: Result Ct -2 0.0 0.0	NO RESUL EndPt	T - REPEAT Analyte Result INVALID INVALID	Probe Check Result PASS PASS	
Analyte R Analyte R Analyte Name SARS-CoV- Flu A 1 Flu A 2	ult: Result Ct -2 0.0 0.0 0.0	NO RESUL EndPt	T - REPEAT Analyte Result INVALID INVALID INVALID	Probe Check Result PASS PASS PASS	
Test Resu Analyte R Analyte Name SARS-CoV- Flu A 1 Flu A 2 Flu B	ult: Result Ct -2 0.0 0.0 0.0 0.0	NO RESUL EndPt	T - REPEAT Analyte Result INVALID INVALID INVALID INVALID	Probe Check Result PASS PASS PASS PASS	
Test Resu Analyte R Analyte Name SARS-CoV- Flu A 1 Flu A 2 Flu B RSV	ult: Result Ct -2 0.0 0.0 0.0 0.0 0.0 0.0	NO RESUL EndPt 1 0 2 -4 -2	T - REPEAT Analyte Result INVALID INVALID INVALID INVALID INVALID	Probe Check Result PASS PASS PASS PASS PASS PASS	

Possible causes

- Improper sample collection of preparation
- Presence of interfering substances in the sample

Solution

Repeat the test with a new cartridge



NO RESULT - REPEAT TEST

ACK		HOME	RESULTS	QC	ADMIN
Test Failed			UPLOAD		REPORT
Module A1		Result	Uploa	aded: No	
Sample ID	220155923501	NO RESULT - REP	EAT TEST		
Patient ID					
Test Type	Specimen				
Assay Name	Xpress SARS-CoV-2_Flu_RSV plus				
User	Admin1				
Start Date & Time	01/25/22 08:30:40				
Test Disclaimer	For In Vitro Diagnostic Use Only. For use under the Emergency Use Authorization (US). Test Methodology: RT- PCR				

Presence or absence of the target RNAs cannot be determined.

- SARS-CoV-2: NO RESULT
 SPC: NO RESULT
- Flu A: NO RESULT
- Flu B: NO RESULT
- RSV: NO RESULT

If the probe check passes, the error is caused by the maximum pressure limit exceeding the acceptable range or by a system component failure.

			Test	Report	
Patient ID: Sample ID* Test Type:	:	22015 Speci	5923501 men		
Assay Infor	mation				
Assay Nam	e			Assay Version	Assay Type
Xpress SARS	-CoV-2_F	lu_RSV plus		1	In Vitro Diagnostic
Test Result	:	NO RESUL	T-REPEAT T	EST	
Test Result Analyte Res	: sult	NO RESUL	T-REPEAT T	EST	
Test Result Analyte Res Analyte Name	: sult Ct	NO RESUL	T-REPEAT T Analyte Result	EST Probe Check	
Test Result Analyte Res Analyte Name	: sult Ct	NO RESUL	T-REPEAT T Analyte Result	EST Probe Check Result	
Test Result Analyte Res Analyte Name SARS-CoV-2	sult Ct 0.0	NO RESUL	T-REPEAT T Analyte Result NO RESULT	EST Probe Check Result PASS	
Test Result Analyte Res Analyte Name SARS-CoV-2 Flu A 1	: sult Ct 0.0 0.0	NO RESUL EndPt	T-REPEAT T Analyte Result NO RESULT NO RESULT	Probe Check Result PASS PASS	
Test Result Analyte Re Analyte Name SARS-CoV-2 Flu A 1 FluA2	: Sult Ct 0.0 0.0 0.0	NO RESUL EndPt	T-REPEAT T Analyte Result NO RESULT NO RESULT NO RESULT	Probe Check Result PASS PASS PASS	
Test Result Analyte Re Analyte Name SARS-CoV-2 Flu A 1 FluA2 Flu B	: Ct 0.0 0.0 0.0 0.0 0.0	NO RESUL EndPt	T-REPEAT T Analyte Result NO RESULT NO RESULT NO RESULT NO RESULT	Probe Check Result PASS PASS PASS FAIL	
Test Result Analyte Re Analyte Name SARS-CoV-2 Flu A 1 FluA2 Flu B RSV	: Ct 0.0 0.0 0.0 0.0 0.0 0.0 0.0	NO RESUL EndPt	T-REPEAT T Analyte Result NO RESULT NO RESULT NO RESULT NO RESULT NO RESULT	Probe Check Result PASS PASS PASS FAIL PASS	

Solution

· Repeat the test with a new cartridge



Probe Check: FAIL; all or one

of the probe check results fail

INSTRUMENT ERROR

BACK		HOME	RESULTS	QC	ADMIN	
Test Failed			UPLOAD		REPORT	
Module A2		Result	Uploa	ded: No		
Sample ID	L21120601953	INOTISOMENT EN				
Patient ID						
Test Type	Specimen					
Assay Name	Xpress SARS-CoV-2_Flu_RSV plus					
User						
Start Date & Time	12/06/21 11:43:16					
Test Disclaimer	For In Vitro Diagnostic Use Only. For use under the Emergency Use Authorization (US). Test Methodology: RT- PCR					

Presence or absence of the target RNAs cannot be determined.

• An INSTRUMENT ERROR indicates that insufficient data was collected. For example, the operator stopped a test that was in progress.

			Test	Report	
Patient ID Sample ID Test Type): D*: ::	211206 Specim	601953 nen		
Assay Info	ormation				
Assay Na	me			Assay Version	Assay Type
Xpress SAF	RS-CoV-2_	Flu_RSV plus		1	In Vitro Diagnostic
Analyte R	esult				
Analyte Name	Ct	EndPt	Analyte Result	Probe Check Result	
SARS-CoV	-2 0.0	0	NO RESULT	NA	
Flu A 1	0.0	0	NO RESULT	NA	
Flu A 2	0.0	0	NO RESULT	NA	
Flu B	0.0	0	NO RESULT	NA	
RSV	0.0	0	NO RESULT	NA	
SPC	0.0	0	NO RESULT	NA	

Possible causes

An INSTRUMENT ERROR indicates that insufficient data was collected

- Test was stopped with stop test button; Electrical failure **Solution**
- Secure the power; Repeat the test with a new cartridge



Reasons to Repeat the Test

- A "SARS-Cov-2 PRESOMPTIVE POS" indicates the 2019 novel coronavirus (SARS-CoV-2) nucleic acids may be present. Only one of the SARS-CoV-2 nucleic acid target was detected (E gene) while the other SARS-CoV-2 nucleic acid target (N2 gene) was not detected.
- A "NO RESULT REPEAT TEST" result indicates that the control SPC failed. The sample was not properly processed, PCR is inhibited, or the sample was not properly collected.
- A "NO RESULT REPEAT TEST" result could be due to, but not limited to, Probe Check Control failure, system component failure, no sample added, or the maximum pressure limits were exceeded.
- An "INSTRUMENT ERROR" indicates that insufficient data were collected. For example, cartridge failed integrity test, the operator stopped a test that was in progress, or a power failure occurred.
- If only one viral target is positive but coinfection with multiple targets is suspected, the sample should be re-tested with another FDA cleared, approved, or authorized test, if coinfection would change clinical management.



Technical Assistance

Before contacting Cepheid Technical Support, collect the following GeneXpert[®] information:

Product name	X
Lot number	X
Serial number of the System	X
Software version and, if applicable, Computer Service Tag number	X
Error messages (if any)	X

Log your case online using the following link: http://www.cepheid.com/us/support

 \rightarrow Create a Support Case





Thank You

www.cepheid.com

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